

Actinomycetes: Isolation and Primary Screening for Alkaline Protease Production

Gautam Tanaji Kamble¹, Mukundraj Govindrao Rathod², Shaikh Musveer Mukhtar¹, Sayyad Sahil Anjum¹, Jivan Munja Dhotare³ & Anupama Prabhakarrao Pathak^{1*}

¹ School of Life Sciences (DST-FIST phase-I and UGC-SAP DRS-II sponsored school), Swami Ramanand Teerth Marathwada University, Nanded-431606, Maharashtra, India

² Department of Biotechnology & Bioinformatics (U.G. & P.G.), Yeshwant College of Information Technology, Parbhani 431401 (affiliated to Swami Ramanand Teerth Marathwada University, Nanded), Maharashtra, India

³ College of Agricultural Biotechnology (affiliated to Vasantrao Naik Marathwada Krishi Vidyapeeth, Parbhani), Hatta 431705, Taluka Basmat, Dist. Hingoli, Maharashtra state, India

Email of corresponding author*: anupama.micro@rediffmail.com

Abstract

Actinomycetes are Gram positive and filamentous microorganisms. They are thought to be a transition group between bacteria and fungi. Actinomycetes are widely distributed in different habitats such as fresh and marine water, agricultural and sediment soil and mangrove habitats. They are able to secrete industrially important extracellular multi-catalytic enzymes such as amylases lipases and proteases. Among the different classes of proteases, alkaline proteases are attracting attention of researchers and industrialists due to their special robust properties and applications. In present investigation, we had collected soil samples from local slaughter house of Nanded and isolated efficient alkaline protease producer actinomycete and designated as MK3. It was Gram positive long filamentous rod with many branches and spores. It was catalase, amylase, protease, and lipase positive. This isolate could be exploited in further research for production, purification and characterization of extracellular alkaline protease and could be used in different biotechnological applications.

Keywords: Alkaline Protease Producer, Actinomycetes, Alkaline Soil, Protease Screening, Slaughter House

1. Introduction

Actinomycetes are Gram positive and filamentous microorganisms. They belong to the phylum actinobacteria. They are thought to be a transition group between bacteria and fungi. Actinomycetes are widely distributed in nature. They are well known for their beneficial properties such as production and secretion of extracellular multi-catalytic enzymes, antimicrobial compounds, some specific secondary metabolites etc. In several studies, production of antibacterial and antifungal compounds from actinomycetes has been reported [1]. Moreover actinomycetes have been extensively studied under the broad area of biocontrol of plant diseases. Thermophilic actinomycetes are well known components of the microflora of composts [2, 3]. Composts are mainly for cultivation of button mushroom [4,5]. Soil actinomycetes act as potential source of biopesticides [6, 7]. Selected species of actinomycetes have applications in increased methanogenesis and odor control [8, 9]. Recently some important inferences about actinobacterial metabolites to combat Corona virus are known [10,11].

Proteolytic enzymes play a specific catalytic role in the hydrolysis of proteins. They are widespread in all living organisms as they are essential for cell growth and differentiation. Alkaline proteases are a physiologically and commercially important group of enzymes used in various branches of industries such as the detergent, food, leather tanning, pharmaceutical, and textile industries [12-21]. Many microorganisms produce proteases; however, the enormous extent of commercial alkaline proteases is generally obtained from *Bacillus* sp., as a result of its capacity to secrete large amount of alkaline proteases having significant proteolytic activity and stability at a wide range of pH. Number of studies has been reported on isolation, screening, production and applications of alkaline proteases. Alkaline proteases are attracting attention of researchers due to their special properties and applications [21-29].

Natural weathering of the soil particles, irrigation and flood are some causes of changing pH of the soil. Alkalinity of soil is associated with the presence of sodium carbonate and sodium bicarbonate in the soil. The soil to the close vicinity of slaughter house tends to have a diverse microbial population since soil of this area is often mixed with animal wastes.

Detergents are used to perform washing practices of slaughtering equipments and as such water mixes with the soil there, the pH of the soil is likely to be alkaline [30-32]. Hence in present study, we collected soil samples from local slaughter house of Nanded and five cultivable actinomycetes isolated. These isolates were screened for their proteolytic activity and an efficient alkaline protease producing isolate was characterized.

2. Materials and Methods

2.1. Soil sample collection

Soil samples were collected from local slaughter house, near Degluar naka region of Nanded, Maharashtra, India. The samples were collected in sterilized cotton bags and transported to the laboratory within 12 hours. 1 gm of composite soil sample was moistened with few drops of distilled water and pH was recorded by pH paper strips (HiMedia, Mumbai).

2.2. Isolation of Actinomycetes

One gram of this soil sample was transferred to a flask containing 100 ml sterilized distilled water and kept on rotary shaker at 30°C and 120 rpm for 15 min. Serial dilutions from 10^{-1} to 10^{-10} were prepared and 0.1 ml of each dilution was inoculated and spread on Actinomycetes isolation agar plates (Hi media, Mumbai). The plates were incubated for 6 days at 30°C and characteristics of morphologically different colonies appearing on the medium were isolated [36-39].

2.3. Screening of selected isolates for proteolytic activity

The selected colonies from Actinomycetes isolation agar plate were screened for proteolytic activity. In this screening method, selected colonies were spot inoculated on alkaline skimmed milk agar plates (pH 8) and incubated at 30°C for 24 to 72 h [40-47].

2.4. Biochemical characterization

For the preliminary characterization of the selected promising isolate the biochemical tests were performed such as IMViC test, catalase production and hydrolysis of lipid, casein, starch, cellulose and urea. Carbohydrate fermentation tests were performed by using standard procedures [48-64].

3. Results and Discussion

The recorded pH of composite soil sample was near about 8.0. This indicated that, the collected soil sample was alkaline. Total 91 colonies were appeared in an average on actinomycetes agar plates, out of which 5 colonies were designated as MK1 to MK5. These colonies were sub-cultured on actinomycetes agar slants. Pure cultures of these isolates were obtained with great difficulty by performing repeated sub-culturing technique. Out of 5 isolates, the isolate MK3 has shown a zone of clearance around it's colony on alkaline skimmed milk agar plate (Figure 1). Morphological and microscopic characters of MK3 isolate are given in Table 1. The observed colony characters, microscopic filamentous view and cellular arrangement resembles that the MK3 belongs to actinomycetes group when compared to reference strain. Biochemical characters of MK3 isolate are given in Table 2.

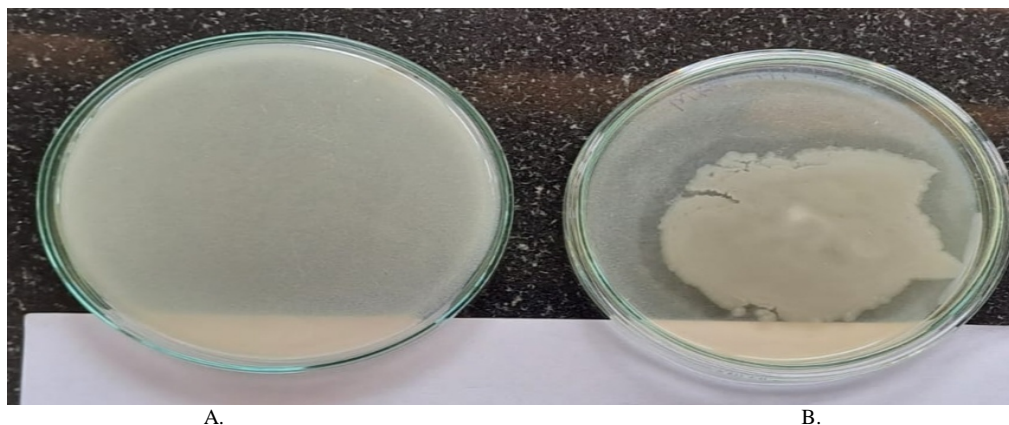


Fig. 1: Primary screening of Actinomycetes for extracellular alkaline protease secretion.

A: Skimmed milk agar plate (pH 8); B: Zone of clearance around the colony of actinomycetes on skimmed milk agar plate (pH 8)

Table 1: Morphological and microscopic characters of MK3 isolate

<i>Character</i>	<i>MK3 isolate</i>
Size	3 mm
Shape	Circular
Color	Off white
Margin	Irregular
Elevation	Raised
Surface	Smooth
Consistency	Sticky
Opacity	Opaque
Gram stain reaction	Gram Positive
Growth of colonies	Highly spreading

Table 2: Biochemical characters of MK3 isolate

Test	Result	Enzyme profile	Results	Carbohydrate fermentation test	Results
Catalase	+	Protease	+	Lactose	+
Indole production	-	Urease	-	Maltose	+
Methyl red	-	Amylase	+	Galactose	+
VP test	-	Lipase	+	Sucrose	+
Citrate utilization	+	Pectinase	+	Inulin	+
Nitrate reduction	-	cellulase	-	Raffinose	-
				Xylose	-
				Trehalose	+
				Melibiose	-
				Salicine	-

In the literature, isolation of actinomycetes has been reported from different sources such as humus layer of forest soil, corn field, cow barn yard, Antarctic Soil, Mitidja plain (Algeria), marine soil, stream sediments and lake muds, marine sediments, root and stem samples of *Cinnamomum zeylanicum*, *Zingiber spectabile*, *Elettariopsis curtisii* and *Labisia pumila*, mangroove sediments etc. However the most common source for the isolation of actinomycetes is agricultural soil sample. Malviya et al. (2013) have isolated actinomycetes from specific soil samples collected after fire operations at agricultural sites under shifting cultivation in northeast India [65]. Many actinomyces species are opportunistic pathogens. Actinobacteria also play an important role as symbionts and as pathogens in plant-associated microbial communities.

Researchers have used different media for isolation of actinomycetes such as starch-casein medium, humic acid-vitamin agar, starch casein nitrate agar (SCS), hair hydrolysate vitamin agar (HHVA), Bennet’s agar (BA), arginine-glycerol salt (AGS) medium, chitin medium, modified Benedict’s medium, soybean meal-glucose medium, Gauze’s agar medium, Czapek’s agar medium, egg albumen medium, glucose-asparagine medium, glycerol-asparaginate agar, chitin agar, coal-vitamin agar, mineral salt (MS) medium, yeast extract-malt extract agar, starch casein nitrate (SCN) agar medium, M3 agar medium, asparagine agar, glycerol-glycine agar, AIM medium, Actinomycetes isolation agar (AIA), tap water yeast extract agar (TWYE) and asparagine-glucose agar medium. The most common medium for cultivation of actinomycetes is starch-yeast extract agar [1,6,8,10,65].

4. Conclusion

The isolate MK3 has shown morphological, microscopic and biochemical characters almost same as with standard reference strains of actinomycetes. Hence it has been concluded that the isolate MK3 belongs to actinomycetes. MK3 has shown proteolytic zone of clearance around its colony on skimmed milk agar plate (pH 8). This isolate could be exploited in further research for production, purification and characterization of extracellular alkaline protease.

Conflict of Interest None.

Acknowledgement

We are thankful to Hon. Dr. Udhav V. Bhosle, Vice-Chancellor of Swami Ramanand Teerth Marathwada University, Nanded for providing infrastructure and necessary facilities to perform research work. Mr. Gautam Kamble highly acknowledges Dr. Babasaheb Ambedkar National Research Fellowship (BANRF 2019) for financial assistance in the form of JRF and SRF.

References

- [1] Sharma M., Dangi, P., & Choudhary, M. (2014). Actinomycetes: source, identification, and their applications. *International Journal of Current Microbiology and Applied Sciences*, 3(2), 801-832.
- [2] Pathak AP, Rathod MG, Akolkar DU, Pansare RR, Sonawane KS, Rathod KK (2021) Microbial biofertilizers for agricultural sustainability In: *Frontiers in Life Science (Volume IV)* ISBN: 978-81-953600-5-5 4, 49-57 Bhumi Publishing, Nigave Khalasa, Kolhapur 416207, Maharashtra
- [3] Pathak AP, Rathod MG, Cherekar MN, Sarsar MS (2022) *Methods in microbiology of extremophiles Volume I* ISBN: 978-93-91120-01-6. Publisher: Akshita Publishers and distributors. J-180, Ram Pratap Marg, new Delhi.
- [4] Rathod MG., Gadade RB, Thakur GM, and Pathak AP. (2021) Oyster mushroom: cultivation, bioactive significance and commercial status. In: *Frontiers in Life Science (Volume II)*: 21. Bhumi publishing, Kolhapur, India.
- [5] Pathak AP, Rathod MG. (2022) *Biotechnology of Mushroom*. 1-69. Bhumi publishing, India ISBN: 978-93-91768-86-7
- [6] Roychoudhury N., & Joshi K.C. (2007) Soil actinomycetes: a potential source of biopesticides. *Vaniki Sandesh* 31(2) 1-6.
- [7] Pathak AP, Rathod MG, Devarshe AM, Hundekar MR, Tengse SA, Kamble GT (2021) Entomopathogenic microorganisms as biopesticides: a review. In: *Frontiers in Life Science (Volume III)* ISBN: 978-81-953600-3-1 3, 95-100 Bhumi Publishing, India
- [8] Duran, M., Tepe, N., Yurtsever, D., Punzi, V. L., Bruno, C., & Mehta, R. J. (2006). Bioaugmenting anaerobic digestion of biosolids with selected strains of *Bacillus*, *Pseudomonas*, and *Actinomycetes* species for increased methanogenesis and odor control. *Applied microbiology and biotechnology*, 73(4), 960-966.
- [9] Pathak AP, Rathod MG, Gaikwad AN *Biology and significance of methanogens*. (2021) In: *Frontiers in Life Science (Volume I)* ISBN: 978-81-951982-1-4 1, 183-188 Bhumi Publishing, India.
- [10] Manikkam, R., Parthasarathy, K., Baskaran, A., & Dellibabu, L. (2022). Inferences of actinobacterial metabolites to combat Corona virus. *Advances in Traditional Medicine*, 1-8.
- [11] Pathak AP, Rathod MG, Ghongade SJ, Katekar PG (2021^d) A scenario on novel corona virus disease (covid-19) pandemic. In: *COVID 19: Impact and Response (Volume II)* ISBN: 978-81-951982-8-3 2, 171-178 Bhumi Publishing, India.
- [12] Rathod MG, Pathak AP (2016^b) Production, extraction and characterization of cold active and salt stable alkaline protease from *Halomonas* sp. LAP520: Lonar soda lake isolate *International Journal of Advanced Research and Review* 1 (5), 123-127
- [13] Rathod, M.G., & Pathak, A.P. (2016). Data on optimized production and characterization of alkaline proteases from newly isolated alkaliphiles from Lonar soda lake, India. *Data in brief*, 8, 863-866.
- [14] Rathod MG, Pathak AP (2017) Evaluation of the enzymatic profile of microbial isolates from Lonar Soda Lake *Indian Journal of Geo-Marine Sciences* 46 (1), 116-124
- [15] Rathod MG, Pathak AP (2014) Isolation and Identification of Alkaline Protease Producer from Selected Alkaline Habitat *Int. J. Innov. Biol. Res.* 3 (1), 1-6; DOI: 10.1111/ijibr.v
- [16] Rathod MG, Pathak AP (2019) Lonar soda lake: a natural habitat of the industrially important diverse alkaline protease producers *Research & Reviews: A Journal of Life Sciences* 9 (1), 81-93
- [17] Rathod M. G., & Pathak, A. P. (2018). Efficient decolorization of textile dyes by alkaline protease producing bacterial consortia. *Indian J. Mar. Sci.* 47(7) 1468-1477.
- [18] Rathod M.G., & Pathak A.P. (2014). Wealth from waste: Optimized alkaline protease production from agro-industrial residues by *Bacillus alcalophilus* LW8 and its biotechnological applications. *Journal of Taibah University for Science*, 8(4), 307-314.
- [19] Rathod M.G., & Pathak, A.P. (2016^c). Optimized production, characterization and application of alkaline proteases from taxonomically assessed microbial isolates from Lonar soda lake, India. *Biocatalysis and agricultural biotechnology*, 7, 164-173.
- [20] Pathak, A.P., & Rathod, M.G. (2021). Exploration of a hot spring for thermostable protease producers. *Journal of Microbiology, Biotechnology and Food Sciences*, 2021, 101-109.

- [21] Pathak AP, Rathod MG, Kamble GT, Basve DM, Mahalinge SV, Ankamwar HG, Chavan S, Mane VS, Thakur GM, Dhopate S, Khule G (2020) Biotechnological Prospective of a Group of Beneficial Extremophiles Isolated from Different Sources Research & Reviews: A Journal of Biotechnology 10 (2), 10-13
- [22] Pathak AP, Rathod MG (2013^b) Utilization of agricultural residues for production of alkaline protease by alkaliphilic *Bacillus firmus* LAP52 : A Lonar soda lake isolate. National conference on Emerging trends in plant sciences (ETPS-2013), 14-19
- [23] Pathak A. P. & Rathod M. G. (2013^a). Production and characterization of alkaline protease by *Bacillus pasteurii*: a Lonar soda lake isolate. *Innov. Res. Chem*, 1(1), 22-26.
- [24] Pathak A. P., Rathod, M. G., Mahabole, M. P., & Khairnar, R. S. (2020). Enhanced catalytic activity of *Bacillus aryabhattai* P1 protease by modulation with nanoactivator. *Heliyon*, 6(6), e04053.
- [25] Pathak A.P., & Sardar, A.G. (2014). Isolation and characterization of salt stable protease producing archaea from marine solar saltern of Mulund, Mumbai.
- [26] Pathak AP, Rathod MG. (2015) A report on thermostable alkaline protease producing bacteria from a terrestrial thermal spring *Indian J. Mar. Sci.* 44 (7), 1104-1111
- [27] Pathak AP, Rathod MG. (2018) A review on alkaline protease producers and their biotechnological perspectives *Indian J. Mar. Sci.* 47 (6), 1113-1119
- [28] Pathak AP, Rathod MG. (2020) Alkaliphilic Microbes: Distribution, Diversity and Applications *International Journal of Industrial Biotechnology and Biomaterials* 5 (2), 32-36
- [29] Pathak AP, Rathod MG (2017) Production and characterization of thermostable gelatinase from *Bacillus globisporus* isolated from Unkeshwar hot spring of Maharashtra *Indian J. Mar. Sci.* 46 (9), 1883-1888
- [30] Ashturkar AR, Behere TR, Bele RB, Rathod MG, Pathak AP (2019) Efficacy of newly formulated inexpensive herbal handwash against common opportunistic pathogens. *Ajanta* 8 (1), 15-18
- [31] Cherekar M. N., & Pathak, A. P. (2015). Studies on haloalkaliphilic gammaproteobacteria from hypersaline Sambhar Lake, Rajasthan, India. *Indian J. Mar. Sci.* 44(10):1646-1653
- [32] Cherekar, M.N., & Pathak, A.P. (2016). Chemical assessment of Sambhar Soda lake, a Ramsar site in India. *Journal of Water Chemistry and Technology*, 38(4), 244-247.
- [33] Girdhari S., & Pathak, A. (2022^a). Thermophilic microbes present in coastal region of Maharashtra. *BIOINFOLET-A Quarterly Journal of Life Sciences*, 19(2), 124-124.
- [34] Girdhari S., & Pathak, A. P. (2021). Application of enzyme extracts of bacterial pectinase and mannanase in fabric stain removal. *Journal of Advanced Scientific Research*, 2021(HBIA), 59-63.
- [35] Girdhari S., & Pathak, A. P. (2022^b). Isolation and identification of thermo stable multi catalytic *Bacillus licheniformis* strain V7 from Ganeshpuri hot spring. *Indian Journal of Natural Products and Resources (IJNPR)[Formerly Natural Product Radiance (NPR)]*, 13(3), 356-361.
- [36] Hingole S. S. & Pathak A. P. (2016^a). Saline soil microbiome: A rich source of halotolerant PGPR. *Journal of crop Science and biotechnology*, 19(3), 231-239.
- [37] Hingole S.S., & Pathak, A.P. (2013). Report on efficient salt stable *Azospirillum* a Lonar Soda Lake isolate. *Science Research Reporter*, 3(2), 200-203.
- [38] Hingole, S.S., & Pathak, A.P. (2016^b). Isolation of halotolerant Plant growth promoting *Klebsiella pneumoniae* from Tuppa, Nanded, Maharashtra. *Int. J. Innov. Biol. Res*, 5(6).
- [39] Jadhav, S.R., & Pathak, A.P. (2019). Production and characterization of a thermo-pH stable pectinase from *Bacillus licheniformis* UNP-1: a novel strain isolated from Unapdev hot spring. *Indian J. Mar. Sci.* 48(5) 670-677.
- [40] Pathak A. P., & Cherekar, M. N. (2015). Hydrobiology of hypersaline Sambhar salt Lake a Ramsar site, Rajasthan, India.
- [41] Pathak A.P., Devkate, S.B., & Rathod, M.G. (2016^a). Production of amylase by *Penicillium* sp. using solid state fermentation method and inexpensive agricultural residues. *International Journal For Research In Biology & Pharmacy*, 2(4), 32-42.
- [42] Pathak A.P., Joshi, V., Murkute, S., & Das, B. (2021). Assessment and determination of halophilic bacterial diversity and antimicrobial potential from mangrove ecosystems of bordi region, maharashtra. *Journal of Advanced Scientific Research*, 2021(HBIA), 71-75.
- [43] Pathak A.P., Sarsar, M.S., Gavali, J.T., & Shendage, M.H. (2016^b). Food and Agricultural residues: Potential substrates for amylase production. *International Journal For Research In Agricultural And Food Science*, 2(4), 01-06.
- [44] Pathak A.P., Sarsar, M.S., Gavali, J.T., & Sonwane, R.G. (2016^c). Amylolytic enzyme production using agricultural residue. *International Journal For Research In Agricultural And Food Science*, 2(4), 30-36.
- [45] Pathak A.P., Sarsar, M.S., Jadhav, S.R., & Kamble, G.T. (2016^d). Extraction, purification and characterization of thermostable lipase from *Thermus* spp.: an industrial effluent isolate. *Int. J. Adv. Res. Rev.* 1(4), 2016; 109-113
- [46] Pathak AP, Khan JF, Marathe KS, Rathod MG (2019) Formulation of efficient herbal fungicide against fungal plant pathogens *Ajanta* 8 (1), 119-123
- [47] Pathak AP, Lodge N, Gavali JT, Rathod MG (2015^a) Isolation and characterization of cold-active protease producer from ice factory samples *Int. J. Adv. Pharm. Biol. Chem. SJIF: 5.548* 4 (4), 751-754
- [48] Pathak AP, Parware SU, Rathod MG (2016^b) Isolation and identification of industrially important salt stable amylase producer *International Journal for Research in Biology & Pharmacy Research* 2 (3), 41-51

- [49] Ujagar AB, Rathod MG, Aasore SR, Pathak AP (2021) Phytochemical screening and antibacterial activity of petroleum ether, methanol and acetone extracts of *Catharanthus roseus* leaves and flowers. In: Research and Development in Pharmaceutical Science (Volume I) 133-141 Bhumi publishing, Kolhapur, India.
- [50] Pathak, A.P., Jethaliya, C.S., Sarsar, M.S., & Jadhav, S.R. (2015^b). Isolation and characterisation of potential amylase producing strain from the agriculture waste. Int. J. Adv. Pharm. Biol. Chem, 4(4), 829-832.
- [51] Pathak, A.P., Kamble, G.T., Jadhav, S.R., & Sarsar, M.S. (2015^c). Isolation and biochemical Characterization of potential thermostable Lipase producer from industrial effluent of oil, dairy and paper industry. Int. J. Adv. Pharm. Biol. Chem, 4(4), 825-828.
- [52] Pathak, A.P., Lohagave, A.G., & Rathod, M.G. (2015^d). Exploration of paper industry effluent for isolation of efficient starchy material degrader to promote bioremediation. Int. J. Adv. Pharm. Biol. Chem, 4(4), 729-736.
- [53] Rathod M.G., Kamble A.D., Shinde P.D., Parsode K.B., Pankhade V.S., Bhosale A.D., Masure N.V., Katkuyare D.M., Pathak A.P. (2019^a) Comparative evaluation of antibacterial activity of honey collected from different trees. Ajanta 8 (1), 115-118
- [54] Rathod MG, Dhembre SC, Thorat SV, Pathak AP (2018) Isolation and Characterization of antibiotic producers from Vajreshwari Hot Spring, Mumbai Bioscience Discovery 9 (4), 474-477
- [55] Rathod MG, Masure N. V., Pankhade V.S., Pathak AP, Shinde PD, Katkuyare DM., Kamble AD, Sonawane SS, Ghatul SK, Bhosale A.D. (2019^b) Biotechnology of microbial polysaccharide: A review. Ajanta 8 (1), 61-65
- [56] Pathak AP, Rathod MG, Rampurkar VV (2014^b) An eco-friendly approach for thermostable amylase production using *Bacillus firmus* APP6: a hot spring isolate Asiatic J Biotech Resources 11, 101-105
- [57] Pathak AP, Rathod MG, Sontakke KT, Kadam TA (2021^c) No vehicle day - A wise step to reduce air pollution: a case study in S.R.T.M. university's campus. In: Ecology Research vol.1, 1-8 Bhumi Publishing, India.
- [58] Pathak AP., Sardar AG, and Janaj PC (2014^b). Exploring the salted fish for salt stable amylase producing bacteria. Indian J. Mar. Sci. 43(10): 1-5.
- [59] Pathak, A.P., & Rathod, M.G. (2014^a). Cultivable bacterial diversity of terrestrial thermal spring of Unkeshwar, India. J. Biochem. Tech, 5, 814-818.
- [60] Pathak, A.P., & Rathod, M.G. (2014^b). Exploration of Unkeshwar hot springs in Maharashtra for thermostable amylase producers. Res. Rev. Biosci, 8(7), 269-276.
- [61] Pathak AP, Rathod MG, Rakhe SA, Bhure NU, Borade RS., Kamalpure PS (2021^e) A step to attain sustainable development in crop production and soil health. In: Ecology Research 2, 1-16
- [62] Pathak AP, Rathod MG, Joshi VB, Girdhari SV (2022^b) Methods in Microbiology of Extremophiles (Vol. II) Methods in Microbiology of Extremophiles 2, 62 Bhumi publishing, Kolhapur, India. ISBN:978-93-91768-20-1
- [63] Pathak AP, Rathod MG, Devarshe AM, Hundekar MR, Tengse SA, Kamble GT (2021^b) Entomopathogenic microorganisms as biopesticides: a review. In: Frontiers in Life Science (Volume III) ISBN: 978-81-953600-3-1 3, 95-100 Bhumi Publishing, India
- [64] Pathak AP, Rathod MG, Akolkar DU, Pansare RR, Sonawane KS, Rathod KK (2021^a) Microbial biofertilizers for agricultural sustainability In: Frontiers in Life Science (Volume IV) ISBN: 978-81-953600-5-5 4, 49-57 Bhumi Publishing, Nigave Khalasa, Kolhapur 416207, Maharashtra
- [65] Malviya, M.K., Pandey, A., Sharma, A., & Tiwari, S. C. (2013). Characterization and identification of actinomycetes isolated from 'fired plots' under shifting cultivation in northeast Himalaya, India. Annals of microbiology, 63(2), 561-569.

Second Author Dr. Mukundraj G. Rathod (Ph.D. in Biotechnology & MH-SET) is in-charge Principal of Yeshwant College of Information Technology, Parbhani, Maharashtra, India. He is also the Head of Biotechnology and Bioinformatics Department of this college. He had published more than 50 research papers including review articles in various peer reviewed International, National journals and proceedings of conferences in different research areas. His Google Scholar citations are 285 with h index & i10 index 9. His Scopus citations are 31 and h index in 04. He had attended many conferences, workshops and presented posters & oral papers. He had also worked as a reviewer for various reputed scientific publishers. He had deposited 7 industrially important cultures at Microbial Culture Collection, National Center for Cell Science, Pune, Maharashtra, India for public use. At present he is the Principal investigator of a research project funded by Swami Ramanand Teerth Marathwada University, Nanded under science and technology application scheme of Rajiv Gandhi Science and Technology Commission, Mumbai, India (Government of Maharashtra). Recently he has been ranked at 17th position amongst the list of top 20 scientists of Swami Ramanand Teerth Marathwada University, Nanded as ranked by AD Scientific Index 2023.

Fifth Author Mr. J.M. Dhotare (B.Sc. Agriculture & M.Sc. Biotechnology) is Asst. Professor at College of Agricultural Biotechnology (affiliated to Vasantrao Naik Marathwada Krishi Vidyapeeth, Parbhani), Hatta, Dist. Hingoli, Maharashtra state, India. He has achieved expertise in teaching r-DNA technology, microbial genetics and related subjects. He has published 2 full length research papers and participated in various extra and co-curricular activities.

Sixth Author Prof. Dr. (Mrs.) Anupam P. Pathak (Ph.D. & MH-SET) is the former Director of School of Life sciences and now Head of Microbiology at School of life sciences of Swami Ramanand Teerth Marathwada University, Nanded. She had published more than 200 research papers including review articles in various peer reviewed International, National journals and proceedings of conferences in different research areas. Her Google Scholar citations are more than 760 with h index 15 & i10 index 28. Her Scopus citations are 200 and h index in 07. She had attended many conferences, workshops and presented posters & oral papers. She had deposited many industrially important bacterial cultures at Microbial Culture Collection, National Center for Cell Science, Pune and more than 110 16S rRNA gene sequences in Genebank, Bethesda, United State of America for public use. She had completed two University Grant Commission, New Delhi funded research projects in Microbiology on extremophiles. She is member of Board of Studies (Microbiology) in this university. She is an eminent scientist and recently she has been ranked at 9th position amongst the list of top 20 scientists of Swami Ramanand Teerth Marathwada University, Nanded as ranked by AD Scientific Index 2023.